

Effect Of Metabotropic Glutamate 5 Receptor Antagonists Modulator In Rats With Neuropathic Pain

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Abstract

The modulation of experimental induced antinociception and antihyperalgesia is mediated by the metabotropic glutamate 5 (mGlu5) receptor, which is also involved in neuropathic pain. Multiple pain models benefit from the antihyperalgesic effects of systemic mGlu5 receptor antagonists (R,S)-alpha-2-methyl-4-sulphonophenylglycine (MSPG). In rats with chronic neuropathic pain, the purpose of this study is to assess the effects of systemic and intrathecal (MSPG) treatment. Rats were used to create a neuropathic pain model by spinal nerve ligation (SNL) at L5–6 and CCI. Behavior tests were used, including the Von Frey and the hot water tail flick tests. On both acute and chronic pain, the effects of low, medium, and high doses of MSPG were investigated. In the Von Frey tests but not the tail flick tests, systemic and intrathecal MSPG both demonstrated dose-dependent antiallodynic effects. The antinociceptive effect in the latter test was actually diminished by high dosages of MSPG. Low dosages of MSPG reduced morphine tolerance in both tests following chronic intrathecal co-administration of MSPG. Only MSPG reduced neuropathic pain systemically, and only in the Von Frey tests—not the tail flick tests—and MSPG had no impact on morphine tolerance in either test. Using mGlu5 receptor antagonists therapeutically may have diverse outcomes in various pain models.

Keywords: NMDA receptor; Rat brain; mGluR1; mGluR5.

1. Introduction

Numerous studies on the antihyperalgesic effects of metabotropic glutamate receptors, in particular subtype 5 receptor, have been conducted in a variety of pain models, including acute and chronic pain including cancer pain, neuropathic pain, visceral pain, and inflammatory pain. MPEP, SIB-1893, and SIB-1757, three noncompetitive mGlu5 receptor antagonists, are currently being employed as effective tools to investigate how endogenous activation of Neurodegeneration and overall neuronal function are impacted by mGlu5 receptors. The metabotropic glutamate 5 (mGlu5) receptor participates in the control of m-opioid-induced antinociception and antihyperalgesia as well as the processing of pain (1,2). The antihyperalgesic effects of the systemic mGlu5 receptor antagonists 2-methyl-6-phenylethynylpyridine (MPEP) and 3-[(2-methyl-1,3-thiazol-4-yl) ethynyl] pyridine (MTEP) is seen in a variety of pain models, but few research have examined their interactions with morphine in models of neuropathic pain (3,4). This study's objective is to find the results of (MSPG) reduced the intensity of these pains when administered systemically, intrathecal administration of MSPG had an antihyperalgesic effect in models of neuropathic pain. Research on the effects of mGlu5 receptor antagonists on neuropathic pain effects in rats (5,6).

2. Materials and methods

2.1. Care for animals

On 25 Wistar rats, the experiments were conducted (150–250 g). The trials were carried out in compliance with the regulations set forth by the institution's animal welfare committees. A 12–12 h light–dark cycle was used for the animal housing process, which was conducted at 23 ± 2 °C, 50 ± 1% relative humidity, and 23 ± 2 °C. Animal care committee approval and compliance with CPCSEA criteria were obtained for experiments involving animals at the School of Pharmacy, Bharat Institutional of Technology Meerut. Before the trial, rats had a 3-day adaptation period. To the randomization and treatments, all investigators were concealed. 06 animals total—three males and three females—were divided equally among each group.

2.2. Experimental Procedures

The 50% paw withdrawal threshold (PWT) was established utilizing the previously stated up-down approach to assess the behavioral response to mechanical stimulation (7–9). Rats were placed in separate plexiglass boxes on a stainless-steel mesh floor and given at least 20 minutes to get used to their new environment. The plantar surface of a hind paw was subjected to a series of calibrated Von Frey monofilaments that were applied perpendicularly and bent for six seconds. Positive reactions were interpreted as quick withdrawals or paw flinching. mGlu5 receptor antagonists as well as to observe their effects (10,11).

2.3. Drugs and Administration

By intrathecal injection, 10% dimethyl sulfoxide (DMSO) served as the vehicle control while mGlu5 receptor antagonist (R,S)-alpha-2-methyl-4-sulphonophenylglycine (MSPG) was used as the antagonist. Drugs were delivered by SC and IT routes after being dissolved in 0.9% saline for the study. On the seventh day following surgery, all medications were started to be administered. A single MSPG dosage was done 30 minutes prior to the saline injection. Following a medication or saline injection, behaviour assessments were conducted 30 minutes later. At 1, 7, 14, 21, and 28 days, behaviour tests were conducted. Behavior tests were conducted every day 30 minutes following the morning therapy. Saline was administered to the control group on the same schedule. Sevoflurane was used to induce and maintain the animals' anaesthesia. Drugs (10,100 g) were then delivered intratracheally at the lumbar 4-5 level using 30-G needles (12).

2.3. Paclitaxel-Induced Neuropathic Pain Model

In a nutshell, paclitaxel was dissolved in a 1:1 (w/v) solution of 0.9% NaCl (normal saline) and 0.2 mg/ml just before administration. Normal saline comprising roughly 1.7% of dosage per dose served as the control substance. Every day, the mice received intraperitoneal (i.p.) injections of paclitaxel (4 mg/kg) or a vehicle control.

3. Psychological Tests

The use of mechanical stimuli, a heat test, and conditioned place preference (CPP) was conducted in accordance with previously published methods (12–15). Animals were taught for three days, spending an hour a day using the testing equipment, to determine baseline threshold and latency. Baseline testing was conducted one day before the introduction of the drug. The two tests were separated by one hour. All behavioural assays were conducted by a single researcher who was blind to all treatments (16).

In response to mechanical stimulation, paw withdrawal frequency (PWF) was initially determined. On the plantar surface of the right hind paws, a 0.4-g von Frey filament was placed. The number of paw withdrawals in response to stimulation was provided as a percent response frequency [(number of paw withdrawals/10 trials) 100]. Stimulations were carried out ten times.

Rodents were separately put in Plexiglass chambers on glass plates and their paw withdrawal latency (PWL), which measures heat sensitivity, was measured by activating the plantar centre with a laser beam. The PWL was calculated as the interval between the initiation of the light beam and the foot withdrawal. In order to prevent tissue damage, tests were run five times at 5-min intervals for a maximum of 20 s.

3.1. Paw Withdrawal Latency (PWL)

Effects of MSPG on paw withdrawal threshold (PWT) and paw withdrawal latency (PWL) in rats following chronic constriction injury (CCI/CINP/DNP). The CCI/CINP/DNP model's experimental setup is shown in the top panel for rats. To study the impact of CCI + MSPG, i.p.) on thermal PWL and mechanical PWT evoked by chronic constriction injury, pentobarbital (40 mg/kg) anaesthesia was administered to rats (CCI). The paw pressure test and heat stimulation were used to measure PWL and PWT before (day -1), as well as on days 1, 3, and 7 and 14 after surgery. Data analysis was done using the GraphPad Prism 9 programme. We created box-and-whiskers plots, where each "box" represented the median, 25th and 75th quartiles, and matching "whisker" represented the 5th and 95th percentiles. With the post-hoc Tukey's test for group comparisons, behavioural data were examined using a two-way repeated measures ANOVA. Based on similar behavioural and biochemical tests, the sample size (n = 6) was determined (17,18). A statistically significant difference was shown by the value of P 0.05.

4. Results

4.1. mGlu5 is the DRG's leading indicator of paclitaxel-Induced Neuropathic Pain.

Groups had similar levels of mechanical allodynia and Von Frey heat sensitivity (n= 6) show in **Figures 1 and 2**. As previously mentioned (19), rats who received paclitaxel intraperitoneally but not the vehicle had thermal hyperalgesia and persistent pain hypersensitivity (markedly elevated PWF after stimulation with a 0.4 g von Frey filament). These side effects began 3 days after the initial paclitaxel therapy and persisted for three weeks or longer (20,21). The behaviour and protein levels in DRG specimens were also time-dependently increased by paclitaxel administration. In comparison to animals that received a vehicle injection, the change in L3-5 DRGs was greater on days 3, 7, 14, 21 and 28 following the initial administration of paclitaxel. Accordingly, on days 3, 7, 14, 21, and 28 compared to the vehicle group, mGluR5 protein levels in L3-5 DRGs rose. As anticipated, the vehicle had no effect on the amount of mGluR5 protein and mRNA in the DRGs at rest. The aforementioned research showed that paclitaxel injection enhanced persistent DRG mGluR5 expression, which may be important for the PINP's initiation and maintenance (22–24).

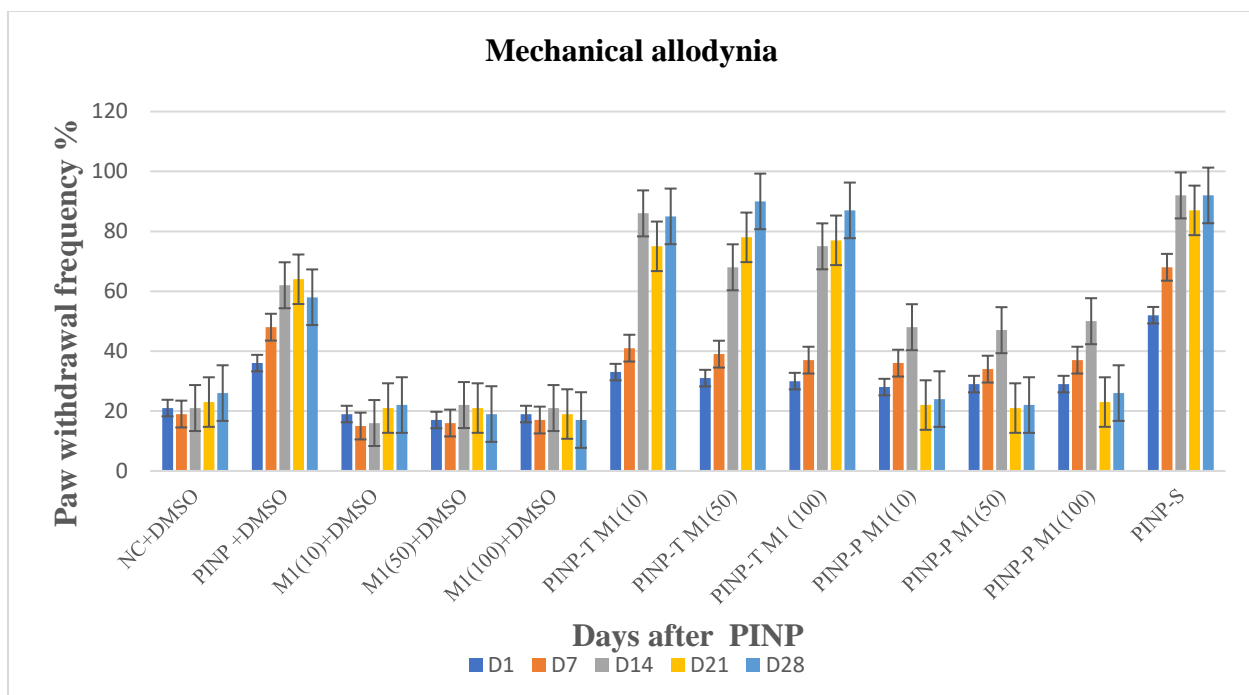


Fig:1 Effects of MSPG on paw withdrawal latency (PWL) and paw withdrawal threshold (PWT) Mechanical allodynia evoked by Paclitaxel-Induced Neuropathic Pain (PINP) in rats.

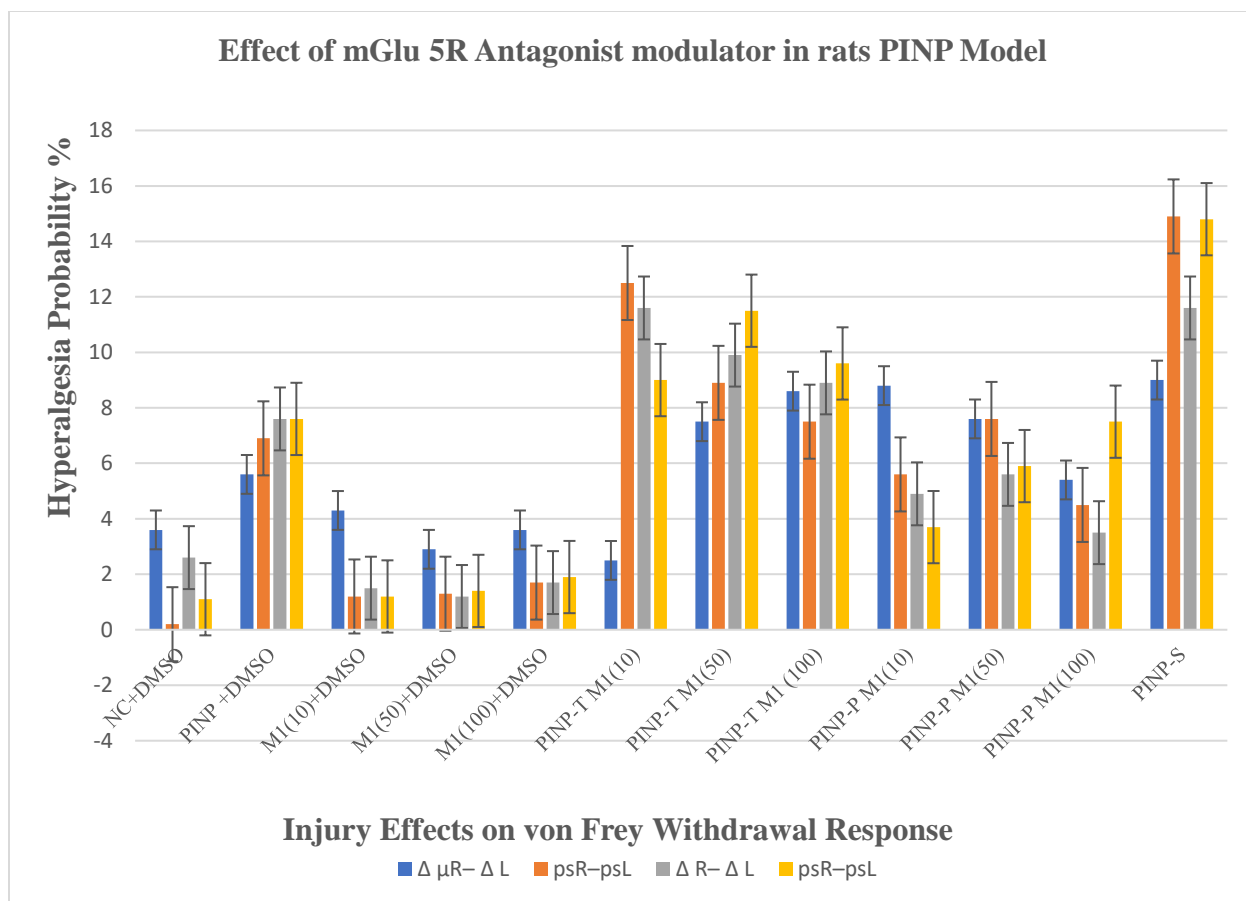
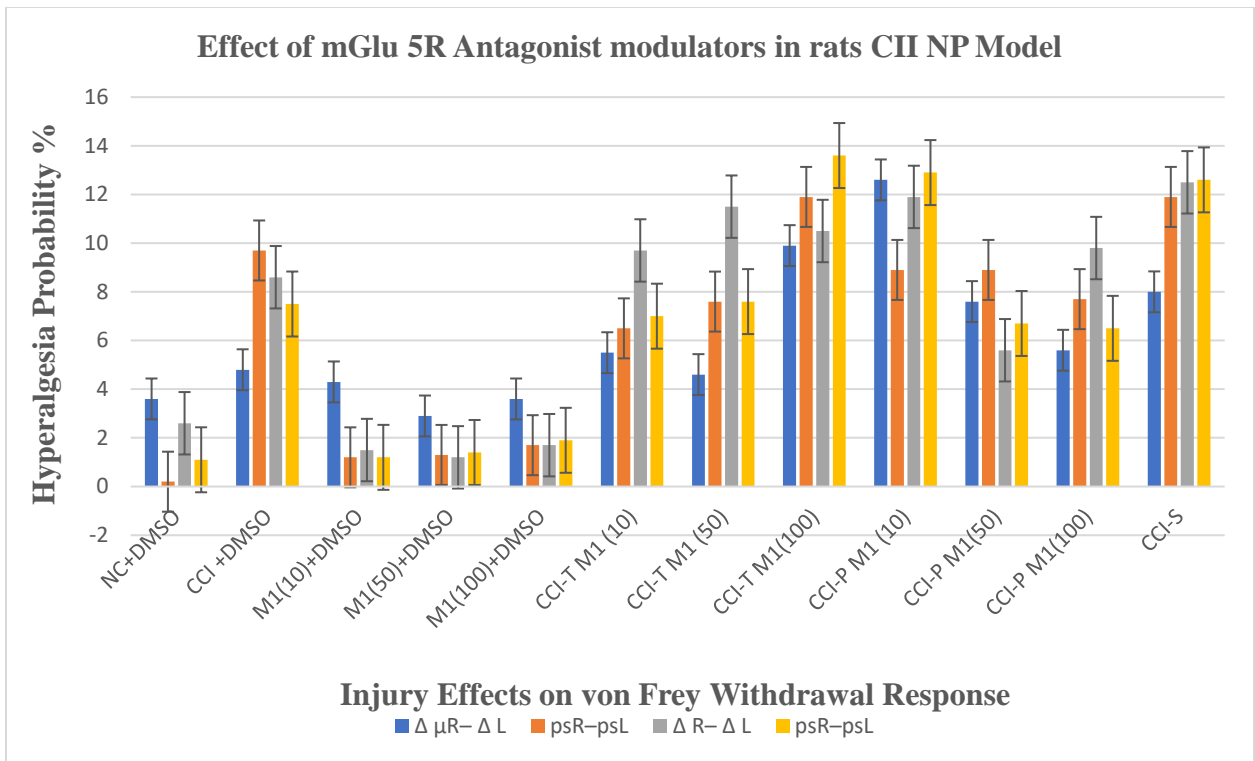


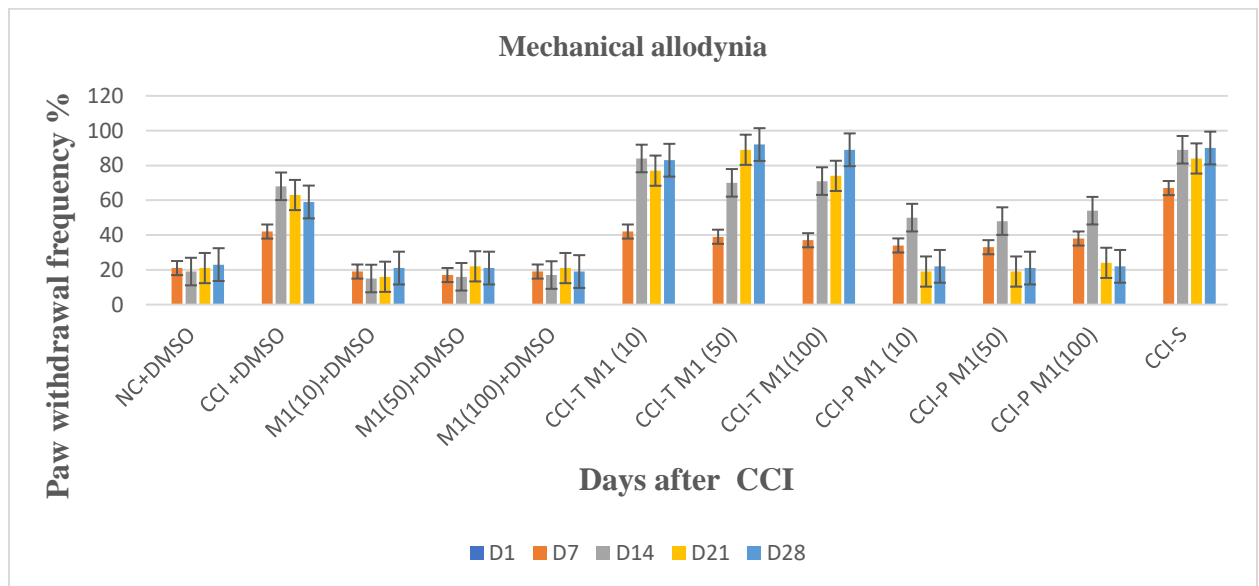
Fig:2 Effects of MSPG on paw withdrawal latency (PWL) and paw withdrawal threshold (PWT) Von Frey evoked by Paclitaxel-Induced Neuropathic Pain (PINP) in rats.

4.2 Chronic constriction injury (CCI) Neuropathic Pain is accompanied mGlu5 in DRG

Mechanical allodynia and Von Frey heat sensitivity were comparable in both groups ($n = 6$) **Figures: 3, Figures:4.** Effects of MSPG on paw withdrawal latency (PWL) and paw withdrawal threshold (PWT) evoked by chronic constriction injury (CCI) in rats. The top panel depicts the experimental design for the CCI model in rats. Rats were anesthetized with pentobarbital (40 mg/kg) for CCI surgery (day 0) to determine the effect of CCI + MSPG, i.p.) on thermal PWL and mechanical PWT evoked by chronic constriction injury (CCI). PWL and PWT were estimated by the thermal stimulation and paw pressure test, respectively, applied before (day -1) and on days 1, 3, 7, 14, 21, 28 after surgery.



Figures 3: Effects of MSPG on paw withdrawal latency (PWL) and paw withdrawal threshold (PWT) Von Frey evoked by chronic constriction injury (CCI) in rats.

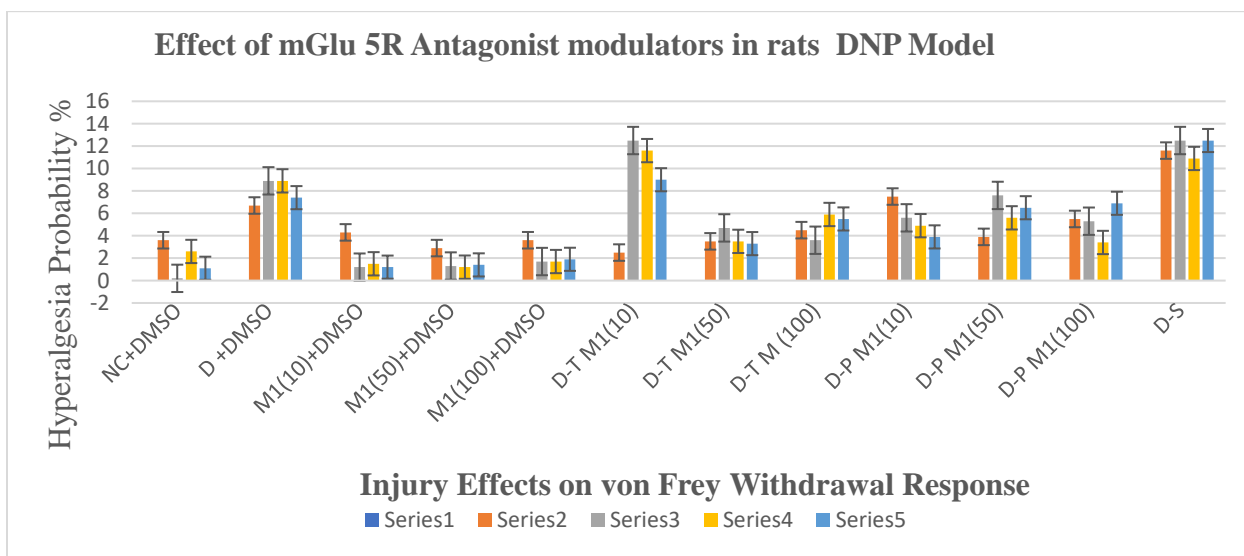


Figures 4: Effects of MSPG on paw withdrawal latency (PWL) and paw withdrawal threshold (PWT) Mechanical allodynia evoked by chronic constriction injury (CCI) Pain in rats.

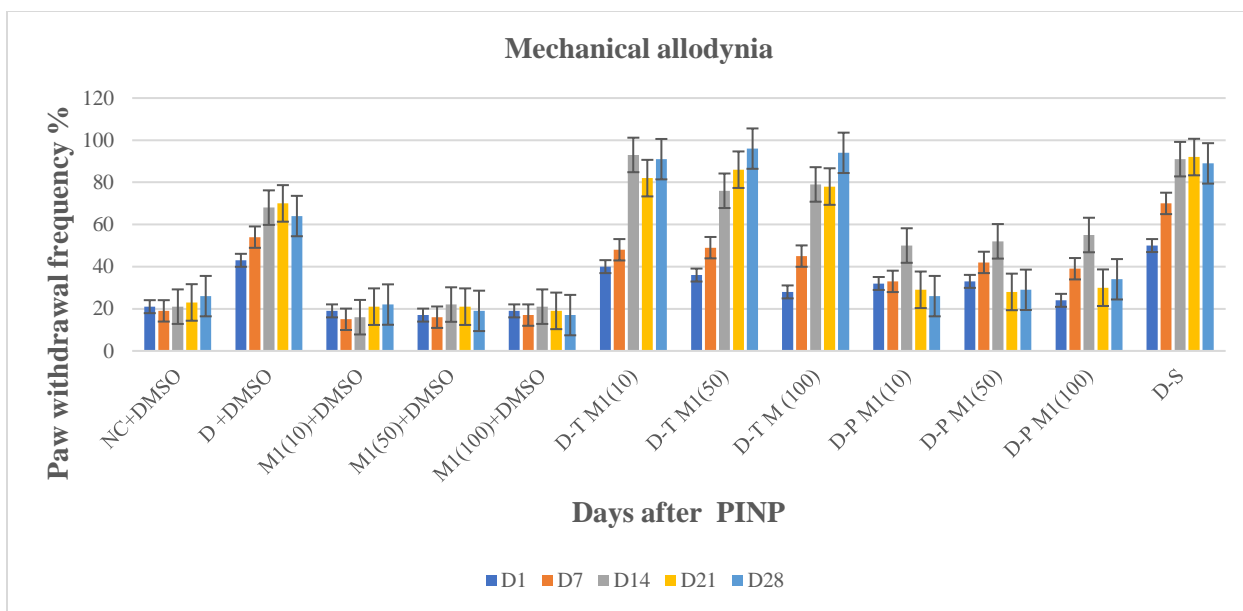
Sham-operated (Sham) rats were subjected to the same surgical procedure, without manipulation of the nerve. Data represent the mean \pm SEM for 6 rats per group. [#]p < 0.05, ^{##}p < 0.01, and ^{###}p < 0.001 compared to the sham group; *p < 0.05 and **p < 0.01 compared to the CCI group.

4.3. Disease-Induced Neuropathic Pain is accompanied mGlu5 Antagonist in DRG

Basal Mechanical allodynia and Von Frey heat sensitivity were comparable in both groups (n = 6) **Figures: 5, Figures:6**. Effects of mGlu5 Antagonist modulators on paw withdrawal latency (PWL) and paw withdrawal threshold (PWT) evoked by diabetic neuropathy. Streptozotocin (STZ)-induced diabetes in the rat has been increasingly used as a model of painful diabetic neuropathy and to assess the efficacies of mGlu 5R antagonist of potential analgesic agents in neuropathic pain. in rats. The top panel depicts the experimental design for to determine the effect of mGlu in Disease-Induced Neuropathic Pain on thermal PWL and mechanical PWT. PWL and PWT were estimated by the thermal stimulation and paw pressure test, respectively, applied before (day -1) and on days 1, 3, 7, 14, 21, 28 after surgery.



Figures 5: Effects of MSPG on paw withdrawal latency (PWL) and paw withdrawal threshold (PWT) Von Frey evoked by Disease-Induced Neuropathic Pain in rats.



Figures 6: Effects of MSPG on paw withdrawal latency (PWL) and paw withdrawal threshold (PWT) Mechanical allodynia evoked by Disease-Induced Neuropathic Pain in rats.

4.4. PINP Is Blunted by Intrathecal MSPG Administration

Intrathecal administration of MSPG (10,50 and 100) was done ten minutes prior to each paclitaxel injection in order to evaluate its specific impact on PINP. First, when compared to regular saline, MSPG had no overt impact on peripheral mechanical and heat sensitivity. It's interesting to note that mice pretreated with MSPG at doses of 10 and 100 mg, but not 1 mg, had lessened the mechanical and thermal allodynia and hyperalgesia caused by paclitaxel, as seen by noticeably reduced PWF control and noticeably increased PWL in the treatment groups. The dose-dependent anti-hyperalgesic effects of MSPG became apparent three days after the initial paclitaxel injection and persisted for 28 days.

4.5. MSPG Inhibits (4.5) Upregulation of mGluR5 by Paclitaxel in DRGs

The quantity of DRG mGluR5 was measured following paclitaxel exposure to determine whether mGluR5 plays a role in the anti-nociceptive impact of MSPG. Unexpectedly, MSPG pretreatment decreased DRG mGluR5 upregulation. According to the aforementioned research, mGluR5 downregulation caused MSPG to impair PINP production and maintenance.

4.6. Inhibition of mGluR5 Lowers PINP

Mice were given intrathecal injections of MSPG (mGluR5 antagonist; 1, 10, or 100 nmol) 10 min before each injection of paclitaxel to validate mGluR5's role in transducing nociception in PINP. As expected, behavioural assays demonstrated that paclitaxel-treated mice had significantly increased paw withdrawal mechanical threshold and thermal latency, indicating that mechanical allodynia and thermal hyperalgesia were robustly and dose-dependently reversed by MSPG at 10 and 100 nmol, but not 1 nmol.

4.7. Blending MSPG in the PINP

Following that, von Frey and hot plate assays showed that MSPG (10, 50,100) and MPEP (1 nmol) at subthreshold doses had no impact when given alone on the paw withdrawal threshold and latency. However, simultaneous

administration of MSPG and MPEP effectively reduced mechanical allodynia and thermal hyperalgesia. The aforementioned results further suggested that mGluR5 was involved in MSPG's anti-PINP activity. In the PINP, MSPG alone has a weaker anti-hyperalgesic action than the combination of the mGluR5 antagonist MPEP and MSPG.

4.8. MSPG's therapeutic effectiveness

Next, it was investigated whether MSPG similarly reduces acute pain brought on by intrathecal administration of the mGluR5 agonist CHPG at 1 nmol. Ten minutes before the CHPG treatment, an intrathecal injection of MSPG (100) was administered. Our findings showed that from 1 to 12 hours after intrathecal dose, CHPG caused acute mechanical allodynia and thermal hyperalgesia in naive rats. It's interesting to note that MSPG medication effectively stopped CHPG-related acute pain. All of the aforementioned information pointed to the critical part played by mGluR5 in anti-nociception brought on by MSPG and chemotherapy-related pain. MSPG Therapy Attenuates Acute Pain Caused by the Metabotropic Glutamate Receptor 5 Agonist 2-Amino-2- (2-Chloro-5-Hydroxyphenyl) Acetic Acid.

4.9. MSPG Controls mGluR5 in DRGs

Ten days after the first dose of paclitaxel was administered, mice's L3-5 DRGs were taken, and the levels of mRNA and membrane protein expression of receptors that are unmistakably linked to pain, such as MOR, NR1, GluA2, and GABAB, were assessed. In paclitaxel + MSPG-treated and paclitaxel + MSPG-treated DRGs, NR1 expression was lowered and GluA2 expression was elevated in comparison to the paclitaxel group.

4.10. MSPG function in Neuroinflammation.

Strong anti-inflammatory properties of MSPG may play a role in PINP's anti-hyperalgesic mechanism. It is currently unknown, nevertheless, whether mGluR5 contributes to the anti-inflammatory action of MSPG. Ten days after the initial paclitaxel therapy, L3-5 DRGs were collected, and chemokines and pro-inflammatory cytokines associated with paclitaxel injection were quantitated. TNF-, IL-1, and IL-6 levels were found to be considerably higher in DRGs treated with paclitaxel, according to our research. As mGluR5 is not implicated in MSPG's anti-inflammatory mechanism, this finding showed that MSPG could lower PINP through an anti-inflammatory mechanism (25,26).

5. Discussion

The inhibition of mGluR5 activation and neuroinflammation by MSPG in the prevention and treatment of PINP was previously unknown, according to this study. From day 3 through at least 28 days following paclitaxel injection, the animals had mechanical allodynia, thermal pain, and spontaneous pain. These symptoms were accompanied by elevated levels of mGluR5, chemokines, and pro-inflammatory substances in DRGs.

A rising number of studies have shown how the development of various neuropsychiatric illnesses is influenced by mGluR5-induced central nervous system (CNS) neurodegeneration. More recently, it has been demonstrated that neurotrauma and peripheral inflammatory reactions activate mGluR5 at excitatory nociceptive synapses, causing chronic neuropathic allodynia and acute inflammatory pain (27,28)

Additionally, animal models are used to control mGluR5-dependent synaptic plasticity in opioid-triggered analgesic tolerance. Here, we describe the mGluR5 expression of mice given paclitaxel for the first time, which supports the progression of the PINP phenotype. The aforementioned data strongly suggests that DRG mGluR5 activation contributes to the pathophysiological process of PINP. However, more research is needed to determine how mGluR5 regulates DRG nociception following paclitaxel treatment.

NMDAR and AMPAR are also co-expressed in neural synapses in addition to mGluR5 (29,30). It is possible that mGluR5 is involved in neuropathic pain because it alters the expression of glutamate receptors in neurons, specifically NMDA (31–33) and AMPA (34) receptors. Presynaptic mGluR5 activity in the spine is increased in paclitaxel-induced neuropathy, acting as an upstream signal for PKC-induced tonic NMDAR induction (35,36). Presynaptic NMDARs are tonically activated by chemotherapy, which encourages glutamate release at synapses in the spinal dorsal horn

(36). Ketamine medication can thereby dramatically ease PINP and lower opioid usage (37). When mGluR5 is activated, neurons produce more NR1 and internalise more GluA2 (38), which results in more NMDAR- and AMPAR-mediated neuronal excitatory afferents. In the situation of spinal nerve ligation neuropathic pain, MOR expression is reduced (39). MMG22, a dual-acting MOR agonist and mGluR5 antagonist, shows strong anti-hyperalgesic effects, especially in the pain associated with cancer, inflammatory pain, and neuropathic pain (40–44). The CNS's major regulators of fast synaptic inhibition are GABAARs (45).

Mechanical allodynia, aberrant neural responses, and dysregulated pro- and anti-inflammatory factors in the spinal dorsal horn are all components of chemotherapy-induced peripheral neurotoxicity (CIPN) (46,47). One of the most important factors in the development of paclitaxel-mediated neuropathic pain is neuroinflammation (48). Strong anti-inflammatory, antioxidant, and neuroprotective properties are demonstrated by MSPG and its derivatives. When BV2 cells are exposed to LPS, artemisinin decreases NO secretion and significantly reduces IL-1, IL-6, and TNF- levels (49).

This study showed that MSPG had excellent therapeutic effects on chemotherapy-related pain and neuropathy. It is important to note that MSPG is inexpensive, low-toxic, and very effective, all of which may contribute to the therapeutic alleviation of CINP. MSPG remarkably crosses the blood-brain barrier effectively (50). MSPG may be well suited for concurrent usage with other chemotherapeutic medicines to avoid the development of chemotherapy pain because it exhibits regulatory effects on several pain-related ion channels, anti-neuroinflammatory activity, and anti-cancer capabilities (51). Chronic inflammation that results from nerve damage fuels the development and maintenance of neuropathic pain. Through their hydrolytic action, inflammatory mediators including secreted phospholipase A2 (sPLA2) control nociceptive and excitatory neural transmission at the onset of pain (52,53). It was shown that the pro-inflammatory mediators in the dorsal root ganglion (DRG) increased in the chemotherapy pain model, indicating the production of neuroinflammation in the DRG. It is generally recognised that trigeminal neuralgia is caused by neuroinflammation in glial cells, which increases presynaptic glutamate release and activates postsynaptic mGluR5 (53,54). An important objective in the management of persistent neuropathic pain should be to balance glutamate release and absorption after nerve injury (55,56). We speculate that neuroinflammation may lead to the release of glutamate, which activates mGluR5, in the pathogenesis of chronic pain. MSPG exerts positive anti-inflammatory effects to reduce the activation of mGluR5, which further results in anti-inflammatory and analgesic effects. This study did not monitor glutamate levels within the DRG, which is one of our limitations.

Additionally, we discovered a really intriguing phenomenon: MSPG significantly increased levels of Myelin Protein Zero (MPZ) (data not shown). Simple MSPG administration could greatly increase MPZ expression. Since MPZ is a crucial protein in Schwann cells and is essential for the production of myelin and the maintenance of proper morphology, it is thought that MSPG greatly upregulates MPZ in DRG (57–59). Additionally, demyelination plays a significant role in the neuropathy caused by paclitaxel (60).

The bottom line is that MSPG can successfully prevent and treat PINP, possibly by preventing mGluR5-related neuroplasticity and lowering neuroinflammation. According to the current findings, MSPG represents a cutting-edge treatment option for neuropathic pain following chemotherapy.

REFERENCE:

1. Kozela E, Wrobel M, Kos T, Wojcikowski J, Daniel WA, Wozniak KM, et al. 2-MPPA, a selective glutamate carboxypeptidase II inhibitor, attenuates morphine tolerance but not dependence in C57/Bl mice. *Psychopharmacology (Berl)* [Internet]. 2005 Dec 12;183(3):275–84. Available from: <http://link.springer.com/10.1007/s00213-005-0182-5>
2. Ren B, Gu X, Zheng Y, Liu C, Wang D, Sun Y, et al. Intrathecal Injection of Metabotropic Glutamate Receptor Subtype 3 and 5 Agonist/Antagonist Attenuates Bone Cancer Pain by Inhibition of Spinal Astrocyte Activation in a Mouse Model. *Anesthesiology* [Internet]. 2012 Jan 1;116(1):122–32. Available from: <https://pubs.asahq.org/anesthesiology/article/116/1/122/11189/Intrathecal-Injection-of-Metabotropic-Glutamate>
3. Kumar N, Laferriere A, Yu JSC, Poon T, Coderre TJ. Metabotropic glutamate receptors (mGluRs) regulate noxious stimulus-induced

- glutamate release in the spinal cord dorsal horn of rats with neuropathic and inflammatory pain. *J Neurochem* [Internet]. 2010 Apr 19;no-no. Available from: <https://onlinelibrary.wiley.com/doi/10.1111/j.1471-4159.2010.06761.x>
4. Urban MO, Hama AT, Bradbury M, Anderson J, Varney MA, Bristow L. Role of metabotropic glutamate receptor subtype 5 (mGluR5) in the maintenance of cold hypersensitivity following a peripheral mononeuropathy in the rat. *Neuropharmacology* [Internet]. 2003 Jun;44(8):983–93. Available from: <https://linkinghub.elsevier.com/retrieve/pii/S0028390803001187>
 5. Zhu X, Li Q, Chang R, Yang D, Song Z, Guo Q, et al. Curcumin alleviates neuropathic pain by inhibiting p300/CBP histone acetyltransferase activity-regulated expression of BDNF and Cox-2 in a rat model. *PLoS One*. 2014;9(3).
 6. Narita M, Suzuki M, Narita M, Niikura K, Nakamura A, Miyatake M, et al. Involvement of spinal metabotropic glutamate receptor 5 in the development of tolerance to morphine-induced antinociception. *J Neurochem* [Internet]. 2005 Sep;94(5):1297–305. Available from: <https://onlinelibrary.wiley.com/doi/10.1111/j.1471-4159.2005.03296.x>
 7. Ho Kim S, Mo Chung J. An experimental model for peripheral neuropathy produced by segmental spinal nerve ligation in the rat. *Pain* [Internet]. 1992 Sep;50(3):355–63. Available from: <https://journals.lww.com/00006396-199209000-00015>
 8. Størkson RV, Kjørsvik A, Tjølsen A, Hole K. Lumbar catheterization of the spinal subarachnoid space in the rat. *J Neurosci Methods* [Internet]. 1996 Apr;65(2):167–72. Available from: <https://linkinghub.elsevier.com/retrieve/pii/0165027095001646>
 9. Chaplan SR, Bach FW, Pogrel JW, Chung JM, Yaksh TL. Quantitative assessment of tactile allodynia in the rat paw. *J Neurosci Methods* [Internet]. 1994 Jul;53(1):55–63. Available from: <https://linkinghub.elsevier.com/retrieve/pii/0165027094901449>
 10. Mao Q, Wu S, Gu X, Du S, Mo K, Sun L, et al. DNMT3a-triggered downregulation of K 2p 1.1 gene in primary sensory neurons contributes to paclitaxel-induced neuropathic pain. *Int J Cancer* [Internet]. 2019 Oct 15;145(8):2122–34. Available from: <https://onlinelibrary.wiley.com/doi/10.1002/ijc.32155>
 11. Qabazard B, Masocha W, Khajah M, Phillips OA. H2S donor GYY4137 ameliorates paclitaxel-induced neuropathic pain in mice. *Biomed Pharmacother*. 2020 Jul 1;127.
 12. Zhang L, Zhao Y, Gao T, Zhang H, Li J, Wang G, et al. MSPG Reduces Remifentanyl-induced Hyperalgesia and Peroxiredoxin-3 Hyperacetylation via Modulating Spinal Metabotropic Glutamate Receptor 5 in Rats. *Neuroscience* [Internet]. 2022 Apr;487:88–98. Available from: <https://linkinghub.elsevier.com/retrieve/pii/S0306452222000033>
 13. Liang L, Gu X, Zhao J-Y, Wu S, Miao X, Xiao J, et al. G9a participates in nerve injury-induced Kcna2 downregulation in primary sensory neurons. *Sci Rep* [Internet]. 2016 Dec 22;6(1):37704. Available from: <http://www.nature.com/articles/srep37704>
 14. Sun L, Zhao J-Y, Gu X, Liang L, Wu S, Mo K, et al. Nerve injury-induced epigenetic silencing of opioid receptors controlled by DNMT3a in primary afferent neurons. *Pain* [Internet]. 2017 Jun 4;158(6):1153–65. Available from: <https://journals.lww.com/00006396-201706000-00022>
 15. Zhao J-Y, Liang L, Gu X, Li Z, Wu S, Sun L, et al. DNA methyltransferase DNMT3a contributes to neuropathic pain by repressing Kcna2 in primary afferent neurons. *Nat Commun* [Internet]. 2017 Mar 8;8(1):14712. Available from: <https://www.nature.com/articles/ncomms14712>
 16. Takeda I, Yoshihara K, Cheung DL, Kobayashi T, Agetsuma M, Tsuda M, et al. Controlled activation of cortical astrocytes modulates neuropathic pain-like behaviour. *Nat Commun* [Internet]. 2022 Jul 14;13(1):4100. Available from: <https://www.nature.com/articles/s41467-022-31773-8>
 17. Notartomaso S, Boccella S, Antenucci N, Ricciardi F, Fazio F, Liberatore F, et al. Analgesic Activity of Cinnabaric Acid in Models of Inflammatory and Neuropathic Pain. *Front Mol Neurosci* [Internet]. 2022 Jun 2;15. Available from: <https://www.frontiersin.org/articles/10.3389/fnmol.2022.892870/full>
 18. Li Y, Guo X, Sun L, Xiao J, Su S, Du S, et al. N 6 -Methyladenosine Demethylase FTO Contributes to Neuropathic Pain by Stabilizing G9a Expression in Primary Sensory Neurons. *Adv Sci* [Internet]. 2020 Jul 27;7(13):1902402. Available from: <https://onlinelibrary.wiley.com/doi/10.1002/advs.201902402>
 19. Luo J, Odaka Y, Huang Z, Cheng B, Liu W, Li L, et al. Dihydroartemisinin Inhibits mTORC1 Signaling by Activating the AMPK Pathway in Rhabdomyosarcoma Tumor Cells. *Cells* [Internet]. 2021 Jun 1;10(6):1363. Available from: <https://www.mdpi.com/2073-4409/10/6/1363>
 20. Xu Y, Jiang Z, Chen X. Mechanisms underlying paclitaxel-induced neuropathic pain: Channels, inflammation and immune regulations. *Eur J Pharmacol* [Internet]. 2022 Oct;933:175288. Available from: <https://linkinghub.elsevier.com/retrieve/pii/S0014299922005490>
 21. Li Y, Yin C, Li X, Liu B, Wang J, Zheng X, et al. Electroacupuncture Alleviates Paclitaxel-Induced Peripheral Neuropathic Pain in Rats

- via Suppressing TLR4 Signaling and TRPV1 Upregulation in Sensory Neurons. *Int J Mol Sci* [Internet]. 2019 Nov 25;20(23):5917. Available from: <https://www.mdpi.com/1422-0067/20/23/5917>
22. Staff NP, Fehrenbacher JC, Caillaud M, Damaj MI, Segal RA, Rieger S. Pathogenesis of paclitaxel-induced peripheral neuropathy: A current review of in vitro and in vivo findings using rodent and human model systems. *Exp Neurol* [Internet]. 2020 Feb;324:113-121. Available from: <https://linkinghub.elsevier.com/retrieve/pii/S0014488619302687>
 23. Li Y, Kang J, Xu Y, Li N, Jiao Y, Wang C, et al. MSPG Alleviates Paclitaxel-Induced Neuropathic Pain in Mice by Decreasing Metabotropic Glutamate Receptor 5 Activity and Neuroinflammation in Primary Sensory Neurons. *Front Mol Neurosci* [Internet]. 2022 May 27;15. Available from: <https://www.frontiersin.org/articles/10.3389/fnmol.2022.902572/full>
 24. Walker K, Reeve A, Bowes M, Winter J, Wotherspoon G, Davis A, et al. mGlu5 receptors and nociceptive function II. mGlu5 receptors functionally expressed on peripheral sensory neurones mediate inflammatory hyperalgesia. *Neuropharmacology* [Internet]. 2001 Jan;40(1):10–9. Available from: <https://linkinghub.elsevier.com/retrieve/pii/S0028390800001143>
 25. Xu L, Hu G, Xing P, Zhou M, Wang D. Paclitaxel alleviates the sepsis-induced acute kidney injury via lnc-MALAT1/miR-370-3p/HMGB1 axis. *Life Sci* [Internet]. 2020 Dec;262:118505. Available from: <https://linkinghub.elsevier.com/retrieve/pii/S0024320520312583>
 26. Al-Mazidi S, Alotaibi M, Nedjadi T, Chaudhary A, Alzoughaibi M, Djouhri L. Blocking of cytokines signalling attenuates evoked and spontaneous neuropathic pain behaviours in the paclitaxel rat model of chemotherapy-induced neuropathy. *Eur J Pain* [Internet]. 2018 Apr;22(4):810–21. Available from: <https://onlinelibrary.wiley.com/doi/10.1002/ejp.1169>
 27. Vincent K, Wang SF, Laferrière A, Kumar N, Coderre TJ. Spinal intracellular metabotropic glutamate receptor 5 (mGluR5) contributes to pain and c-fos expression in a rat model of inflammatory pain. *Pain* [Internet]. 2017 Apr;158(4):705–16. Available from: <https://journals.lww.com/00006396-201704000-00018>
 28. Lai C-Y, Hsieh M-C, Ho Y-C, Wang H-H, Chou D, Wen Y-C, et al. Spinal RNF20-Mediated Histone H2B Monoubiquitylation Regulates mGluR5 Transcription for Neuropathic Allodynia. *J Neurosci* [Internet]. 2018 Oct 24;38(43):9160–74. Available from: <https://www.jneurosci.org/lookup/doi/10.1523/JNEUROSCI.1069-18.2018>
 29. Chen S-R, Zhou H-Y, Byun HS, Pan H-L. Nerve Injury Increases GluA2-Lacking AMPA Receptor Prevalence in Spinal Cords: Functional Significance and Signaling Mechanisms. *J Pharmacol Exp Ther* [Internet]. 2013 Dec;347(3):765–72. Available from: <http://jpet.aspetjournals.org/lookup/doi/10.1124/jpet.113.208363>
 30. Goncalves J, Bartol TM, Camus C, Levet F, Menegolla AP, Sejnowski TJ, et al. Nanoscale co-organization and coactivation of AMPAR, NMDAR, and mGluR at excitatory synapses. *Proc Natl Acad Sci* [Internet]. 2020 Jun 23;117(25):14503–11. Available from: <https://pnas.org/doi/full/10.1073/pnas.1922563117>
 31. Bertaso F, Roussignol G, Worley P, Bockaert J, Fagni L, Ango F. Homer1a-Dependent Crosstalk Between NMDA and Metabotropic Glutamate Receptors in Mouse Neurons. Mei L, editor. *PLoS One* [Internet]. 2010 Mar 18;5(3):e9755. Available from: <https://dx.plos.org/10.1371/journal.pone.0009755>
 32. Dai S-H, Qin N, Chen T, Luo P, Zhang L, Rao W, et al. Activation of mGluR5 Attenuates NMDA-Induced Neurotoxicity through Disruption of the NMDAR-PSD-95 Complex and Preservation of Mitochondrial Function in Differentiated PC12 Cells. *Int J Mol Sci* [Internet]. 2014 Jun 17;15(6):10892–907. Available from: <http://www.mdpi.com/1422-0067/15/6/10892>
 33. Aloisi E, Le Corf K, Dupuis J, Zhang P, Ginger M, Labrousse V, et al. Altered surface mGluR5 dynamics provoke synaptic NMDAR dysfunction and cognitive defects in Fmr1 knockout mice. *Nat Commun* [Internet]. 2017 Oct 24;8(1):1103. Available from: <https://www.nature.com/articles/s41467-017-01191-2>
 34. Huber KM, Gallagher SM, Warren ST, Bear MF. Altered synaptic plasticity in a mouse model of fragile X mental retardation. *Proc Natl Acad Sci* [Internet]. 2002 May 28;99(11):7746–50. Available from: <https://pnas.org/doi/full/10.1073/pnas.122205699>
 35. Zhou M-H, Chen S-R, Wang L, Huang Y, Deng M, Zhang J, et al. Protein Kinase C-Mediated Phosphorylation and $\alpha\delta$ -1 Interdependently Regulate NMDA Receptor Trafficking and Activity. *J Neurosci* [Internet]. 2021 Jul 28;41(30):6415–29. Available from: <https://www.jneurosci.org/lookup/doi/10.1523/JNEUROSCI.0757-21.2021>
 36. Xie J-D, Chen S-R, Pan H-L. Presynaptic mGluR5 receptor controls glutamatergic input through protein kinase C–NMDA receptors in paclitaxel-induced neuropathic pain. *J Biol Chem* [Internet]. 2017 Dec;292(50):20644–54. Available from: <https://linkinghub.elsevier.com/retrieve/pii/S0021925820327964>
 37. Pascual D, Goicoechea C, Burgos E, Martín MI. Antinociceptive effect of three common analgesic drugs on peripheral neuropathy induced by paclitaxel in rats. *Pharmacol Biochem Behav* [Internet]. 2010 May;95(3):331–7. Available from: <https://linkinghub.elsevier.com/retrieve/pii/S0091305710000560>

38. Benneyworth MA, Hearing MC, Asp AJ, Madayag A, Ingebretson AE, Schmidt CE, et al. Synaptic Depotential and mGluR5 Activity in the Nucleus Accumbens Drive Cocaine-Primed Reinstatement of Place Preference. *J Neurosci* [Internet]. 2019 Jun 12;39(24):4785–96. Available from: <https://www.jneurosci.org/lookup/doi/10.1523/JNEUROSCI.3020-17.2019>
39. Wu Q, Wei G, Ji F, Jia S, Wu S, Guo X, et al. TET1 Overexpression Mitigates Neuropathic Pain Through Rescuing the Expression of μ -Opioid Receptor and Kv1.2 in the Primary Sensory Neurons. *Neurotherapeutics* [Internet]. 2019 Apr 4;16(2):491–504. Available from: <http://link.springer.com/10.1007/s13311-018-00689-x>
40. Peterson CD, Kitto KF, Akgün E, Lunzer MM, Riedl MS, Vulchanova L, et al. Bivalent ligand that activates mu opioid receptor and antagonizes mGluR5 receptor reduces neuropathic pain in mice. *Pain* [Internet]. 2017 Dec 1;158(12):2431–41. Available from: <https://journals.lww.com/00006396-201712000-00017>
41. Vincent K, Cornea VM, Jong Y-JI, Laferrière A, Kumar N, Mickeviciute A, et al. Intracellular mGluR5 plays a critical role in neuropathic pain. *Nat Commun* [Internet]. 2016 Apr 3;7(1):10604. Available from: <http://www.nature.com/articles/ncomms10604>
42. Akgün E, Javed MI, Lunzer MM, Smeester BA, Beitz AJ, Portoghese PS. Ligands that interact with putative MOR-mGluR5 heteromer in mice with inflammatory pain produce potent antinociception. *Proc Natl Acad Sci* [Internet]. 2013 Jul 9;110(28):11595–9. Available from: <https://pnas.org/doi/full/10.1073/pnas.1305461110>
43. Shueb SS, Erb SJ, Lunzer MM, Speltz R, Harding-Rose C, Akgün E, et al. Targeting MOR-mGluR5 heteromers reduces bone cancer pain by activating MOR and inhibiting mGluR5. *Neuropharmacology* [Internet]. 2019 Dec;160:107690. Available from: <https://linkinghub.elsevier.com/retrieve/pii/S0028390818308438>
44. Speltz R, Lunzer MM, Shueb SS, Akgün E, Reed R, Kalyuzhny A, et al. The bivalent ligand, MMG22, reduces neuropathic pain after nerve injury without the side effects of traditional opioids. *Pain* [Internet]. 2020 Sep;161(9):2041–57. Available from: <https://journals.lww.com/10.1097/j.pain.0000000000001902>
45. Kiss E, Kins S, Zöller Y, Schilling S, Gorgas K, Groß D, et al. MSPG restores the levels of inhibitory synapse proteins and reduces amyloid- β and C-terminal fragments (CTFs) of the amyloid precursor protein in an AD-mouse model. *Mol Cell Neurosci* [Internet]. 2021 Jun;113:103624. Available from: <https://linkinghub.elsevier.com/retrieve/pii/S1044743121000373>
46. Park SB, Goldstein D, Krishnan A V., Lin CS-Y, Friedlander ML, Cassidy J, et al. Chemotherapy-induced peripheral neurotoxicity: A critical analysis. *CA Cancer J Clin* [Internet]. 2013 Nov;63(6):419–37. Available from: <http://doi.wiley.com/10.3322/caac.21204>
47. Staff NP, Grisold A, Grisold W, Windebank AJ. Chemotherapy-induced peripheral neuropathy: A current review. *Ann Neurol* [Internet]. 2017 Jun;81(6):772–81. Available from: <https://onlinelibrary.wiley.com/doi/10.1002/ana.24951>
48. Meesawatson P, Hathway G, Bennett A, Constantin-Teodosiu D, Chapman V. Spinal neuronal excitability and neuroinflammation in a model of chemotherapeutic neuropathic pain: targeting the resolution pathways. *J Neuroinflammation* [Internet]. 2020 Dec 23;17(1):316. Available from: <https://jneuroinflammation.biomedcentral.com/articles/10.1186/s12974-020-01997-w>
49. Qiang W, Cai W, Yang Q, Yang L, Dai Y, Zhao Z, et al. Artemisinin B Improves Learning and Memory Impairment in AD Dementia Mice by Suppressing Neuroinflammation. *Neuroscience* [Internet]. 2018 Dec;395:1–12. Available from: <https://linkinghub.elsevier.com/retrieve/pii/S0306452218307115>
50. Davis TME, Binh TQ, Ilett KF, Batty KT, Phuông HL, Chiswell GM, et al. Penetration of Dihydroartemisinin into Cerebrospinal Fluid after Administration of Intravenous MSPG in Severe Falciparum Malaria. *Antimicrob Agents Chemother* [Internet]. 2003 Jan;47(1):368–70. Available from: <https://journals.asm.org/doi/10.1128/AAC.47.1.368-370.2003>
51. Crespo-Ortiz MP, Wei MQ. Antitumor Activity of Artemisinin and Its Derivatives: From a Well-Known Antimalarial Agent to a Potential Anticancer Drug. *J Biomed Biotechnol* [Internet]. 2012;2012:1–18. Available from: <http://www.hindawi.com/journals/bmri/2012/247597/>
52. Meade E, Garvey M. The Role of Neuro-Immune Interaction in Chronic Pain Conditions; Functional Somatic Syndrome, Neurogenic Inflammation, and Peripheral Neuropathy. *Int J Mol Sci* [Internet]. 2022 Aug 2;23(15):8574. Available from: <https://www.mdpi.com/1422-0067/23/15/8574>
53. Honda K, Shinoda M, Kondo M, Shimizu K, Yonemoto H, Otsuki K, et al. Sensitization of TRPV1 and TRPA1 via peripheral mGluR5 signaling contributes to thermal and mechanical hypersensitivity. *Pain* [Internet]. 2017 Sep 8;158(9):1754–64. Available from: <https://journals.lww.com/00006396-201709000-00015>
54. Zhang C, Ward J, Dauch JR, Tanzi RE, Cheng HT. Cytokine-mediated inflammation mediates painful neuropathy from metabolic syndrome. McKemy DD, editor. *PLoS One* [Internet]. 2018 Feb 6;13(2):e0192333. Available from: <https://dx.plos.org/10.1371/journal.pone.0192333>

55. Russell J, Anjaneyulu M, Berent-Spilson A. Metabotropic Glutamate Receptors (mGluRs) and Diabetic Neuropathy. *Curr Drug Targets* [Internet]. 2008 Jan 1;9(1):85–93. Available from: <http://www.eurekaselect.com/openurl/content.php?genre=article&issn=1389-4501&volume=9&issue=1&spage=85>
56. Inquimbert P, Bartels K, Babaniyi OB, Barrett LB, Tegeder I, Scholz J. Peripheral nerve injury produces a sustained shift in the balance between glutamate release and uptake in the dorsal horn of the spinal cord. *Pain* [Internet]. 2012 Dec;153(12):2422–31. Available from: <https://journals.lww.com/00006396-201212000-00018>
57. Otani Y, Ohno N, Cui J, Yamaguchi Y, Baba H. Upregulation of large myelin protein zero leads to Charcot–Marie–Tooth disease-like neuropathy in mice. *Commun Biol* [Internet]. 2020 Mar 13;3(1):121. Available from: <https://www.nature.com/articles/s42003-020-0854-z>
58. Torii T, Miyamoto Y, Yamauchi J. Cellular Signal-Regulated Schwann Cell Myelination and Remyelination. In 2019. p. 3–22. Available from: http://link.springer.com/10.1007/978-981-32-9636-7_1
59. Moss KR, Bopp TS, Johnson AE, Höke A. New evidence for secondary axonal degeneration in demyelinating neuropathies. *Neurosci Lett* [Internet]. 2021 Jan;744:135595. Available from: <https://linkinghub.elsevier.com/retrieve/pii/S030439402030865X>
60. Moscarello MA, Mak B, Nguyen TA, Wood DD, Mastronardi F, Ludwin SK. Paclitaxel (Taxol) attenuates clinical disease in a spontaneously demyelinating transgenic mouse and induces remyelination. *Mult Scler J* [Internet]. 2002 Apr 2;8(2):130–8. Available from: <http://journals.sagepub.com/doi/10.1191/1352458502ms776oa>